



**Whole Effluent Toxicity Test Report:  
Washington Beef LLC.**

December 2013

Report date: January 3, 2014

Submitted to:

**Washington Beef LLC.**

201 Elmwood Road  
Toppenish, WA 98948

## 1.0 INTRODUCTION

A whole effluent toxicity test was conducted using effluent samples collected from the Washington Beef LLC wastewater treatment plant in December 2013. A chronic bioassay was conducted using the test organism *Ceriodaphnia dubia* (*Ceriodaphnia*). Testing was performed at Rainier Environmental Laboratory located in Tacoma, Washington.

## 2.0 METHODS

### 2.1 Sample Collection and Transport

Effluent samples were collected into 4-liter (L) LDPE cubitainers by Washington Beef personnel. The samples were packed in coolers containing ice and shipped to Rainier Environmental by overnight delivery service. Appropriate chain-of-custody procedures were employed during collection and transport (Appendix D).

### 2.2 Sample Receipt

Upon arrival at the laboratory, coolers were opened, samples inspected, and the contents verified against information provided on the chain-of-custody forms. Receipt temperature was measured and recorded on the chain-of-custody form. The standard water quality parameters were measured and recorded on sample check-in sheets (Appendix B). Samples were stored at 4°C in the dark until used for testing.

### 2.3 Test Methods

A chronic toxicity test was conducted according to procedures presented by USEPA (2002). The methods are summarized in Table 1. The procedure involved a 7-day static-renewal exposure to the effluent. The endpoints from these tests were *Ceriodaphnia* survival at the end of exposure and reproduction at test termination or production of 3 broods, whichever occurred first. Termination of the test occurred when at least 60 percent of surviving control females produced 3 broods. The test was ended on Day 7.

**Table 1. Summary of methods for the 7-day *Ceriodaphnia* survival and reproduction test.**

Test initiation date and time	12/10/13; 1530h
Test termination date and time	12/17/13; 1500h
Test organism	<i>Ceriodaphnia dubia</i>
Test organism source	In-house cultures
Test organism age	< 24 hours
Test duration	7 days; Test terminated when 60% of controls reached 3 broods
Feeding	1:1 mixture YTC:algal suspension daily
Test chamber; test solution volume	30 mL plastic cup; 15 mL
Test temperature	25 ± 1°C
Dilution water	Diluted mineral water
Test concentrations (% sample)	100, 50, 25, 12.5, 6.25, laboratory control
Number of organisms/chamber	1
Number of replicates	10
Photoperiod	16 hours light/8 hours dark
Aeration	None
Test protocol	EPA-821-R-02-013
Test acceptability criteria for controls	≥ 80% survival; ≥ 15 neonates per surviving adult
Reference toxicant	Sodium chloride

### 3.0 RESULTS AND DISCUSSION

Details of standard water quality measurements conducted upon receipt of samples are provided in Table 2.

**Table 2. Final Effluent sample information.**

Parameter	WET		
Rainier Log-in No.	13-157	13-160	13-168
Collection date and time	12/9/2013; 0720h	12/11/2013; 0725h	12/13/2013; 0740h
Receipt date and time	12/10/2013; 1030h	12/12/2013; 1400h	12/14/2013; 1100h
Receipt temperature (°C)	1.0	2.4	2.4
Dissolved oxygen (mg/L)	6.7	7.0	5.4
pH	7.47	7.35	7.30
Conductivity (µS/cm)	4610	4390	4090
Salinity (ppt)	2.3	2.3	2.0
Hardness (mg/L CaCO <sub>3</sub> )	80	76	76
Alkalinity (mg/L CaCO <sub>3</sub> )	168	160	176
Total Chlorine (mg/L) <sup>a</sup>	<0.03	<0.03	<0.03
Total Ammonia (mg/L) <sup>b</sup>	<1.0	<1.0	<1.0

<sup>a,b</sup> See reference below



Note: Total chlorine and ammonia values are measured by Rainier Environmental to provide additional information in support of the bioassay test procedures. They are not intended to be interpreted as exact values, particularly near the detection limits where interferences are most likely to become apparent.

<sup>a</sup> Total chlorine is measured using a Hach DR/2000 spectrophotometer and colorimetric DPD Total Chlorine Reagent. Under optimum conditions, the method has a range of 0.03 to 2.0 mg/L  $\pm$  0.01 mg/L total chlorine. Compounds in the sample that interfere with chlorine detection include bromine, manganese, chromium, ozone, and peroxides. Additional interferences include extreme pH values and high alkalinity (greater than 300 mg/L Ca CO<sub>3</sub>).

<sup>b</sup> Total ammonia is measured using a Hach DR/2000 spectrophotometer following the salicylate method which uses AmVer Diluent Reagent Test 'N Tube kits. Under optimum conditions, the method has a range of 0.4 to 50.0  $\pm$  0.1 mg/L NH<sub>3</sub>-N. High sample turbidity will give erroneously high values. Additional interferences to the method include extreme pH and high concentrations of magnesium, iron, nitrite, nitrate, or sulfate.

Results for the toxicity tests are summarized in Table 3. Individual statistical summaries for the test and copies of the laboratory bench sheets are provided in the Appendices A-D.

The NOEC (concentration at which no effect on the organisms is detected) was 100 percent sample for survival and 12.5 percent for reproduction. The associated chronic toxicity unit (TUC; 100 percent sample divided by the NOEC) was 1 for survival and 8 for reproduction.

**Table 3. Summary of toxicity test results.**

Sample	Endpoint	NOEC (% effluent)	Chronic Toxicity Unit (TUC) <sup>a</sup>
Final Effluent	Survival	100	1.0
	7-day Reproduction	12.5	8.0

<sup>a</sup> Chronic toxicity unit (TUC = 100  $\div$  NOEC)

#### 4.0 QA/QC

Samples were received in good condition and within the temperature range specified by EPA (2002). The toxicity tests met all acceptability criteria for performance of control organisms. There were no deviations from protocol and water quality parameters remained within the ranges specified in the corresponding test methods throughout the tests.

Results for the most recent reference toxicant test used to monitor laboratory performance and test organism sensitivity are summarized in Table 4 and Appendix C. The coefficients of variation (CV) for the endpoints are also shown in the table. The results for the reference toxicant test fell within the acceptable range of mean  $\pm$  two standard deviations of historical test results indicating that the test organisms were of an appropriate degree of sensitivity.



**Table 4. Reference toxicant test results.**

Species	Endpoint	Date initiated	LC <sub>50</sub> /EC <sub>50</sub>	Acceptable Range	CV (%)
<i>Ceriodaphnia</i>	7d survival	12/10/2013	1.52 g/L NaCl	1.26 - 2.70 g/L	21.1
	7d reproduction	12/10/2013	1.43 g/L NaCl	1.07 - 1.57 g/L	10.1

#### REFERENCES

Tidepool Scientific Software. 2000-2011. CETIS Comprehensive Environmental Toxicity Information System Software, Version 1.8.4.6.

USEPA. 2002. Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms, Fourth Edition. EPA-821-R-02-013. pp. 141-196.

**Appendix A**  
**Statistical Summaries and Raw Bench Sheets**

# CETIS Summary Report

Report Date: 02 Jan-14 13:50 (p 1 of 2)  
Test Code: 1312-039 | 15-2517-8062

## Ceriodaphnia 7-d Survival and Reproduction Test

Rainier Environmental Laboratory

Batch ID: 16-6500-4470	Test Type: Reproduction-Survival (7d)	Analyst: Eric Tollefson
Start Date: 10 Dec-13 15:30	Protocol: EPA/821/R-02-013 (2002)	Diluent: Perrier Water
Ending Date: 17 Dec-13 15:00	Species: Ceriodaphnia dubia	Brine:
Duration: 6d 23h	Source: In-House Culture	Age: <24h
Sample ID: 00-0407-8993	Code: 13-157	Client: Washington Beef
Sample Date: 09 Dec-13 07:20	Material: POTW Effluent	Project:
Receive Date: 10 Dec-13 10:30	Source: Washington Beef (WA0050202)	
Sample Age: 32h (1 °C)	Station: Outfall 002	

## Comparison Summary

Analysis ID	Endpoint	NOEL	LOEL	TOEL	PMSD	TU	Method
06-2978-4992	7d Survival Rate	100	>100	NA	NA	1	Fisher Exact/Bonferroni-Holm Test
21-2554-2975	Reproduction	12.5	25	17.68	36.0%	8	Steel Many-One Rank Sum Test

## Point Estimate Summary

Analysis ID	Endpoint	Level	%	95% LCL	95% UCL	TU	Method
04-9633-9232	7d Survival Rate	LC5	0.4336	0.3023	1.693	230.6	Linear Interpolation (ICPIN)
		LC10	1.055	0.696	12.5	94.77	
		LC15	1.946	1.209	50	51.38	
		LC20	3.224	1.876	63.05	31.02	
		LC25	5.055	2.746	N/A	19.78	
		LC40	100	54.02	N/A	1	
		LC50	>100	N/A	N/A	<1	
13-6887-0457	Reproduction	IC5	0.2596	0.1753	1.289	385.2	Linear Interpolation (ICPIN)
		IC10	0.5867	0.3814	4.239	170.5	
		IC15	0.9986	0.6237	12.3	100.1	
		IC20	1.518	0.9084	13.79	65.9	
		IC25	2.171	1.243	15.31	46.06	
		IC40	5.338	2.642	20.87	18.73	
		IC50	16.05	4.031	52.18	6.232	

## Test Acceptability

Analysis ID	Endpoint	Attribute	Test Stat	TAC Limits	Overlap	Decision
04-9633-9232	7d Survival Rate	Control Resp	1	0.8 - NL	Yes	Passes Acceptability Criteria
06-2978-4992	7d Survival Rate	Control Resp	1	0.8 - NL	Yes	Passes Acceptability Criteria
13-6887-0457	Reproduction	Control Resp	26.1	15 - NL	Yes	Passes Acceptability Criteria
21-2554-2975	Reproduction	Control Resp	26.1	15 - NL	Yes	Passes Acceptability Criteria
21-2554-2975	Reproduction	PMSD	0.3603	0.13 - 0.47	Yes	Passes Acceptability Criteria

## 7d Survival Rate Summary

C-%	Control Type	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	Dilution Water	10	1	1	1	1	1	0	0	0.0%	0.0%
6.25		10	0.5	0.3032	0.6968	0	1	0.1667	0.527	105.4%	50.0%
12.5		10	0.8	0.6426	0.9574	0	1	0.1333	0.4216	52.7%	20.0%
25		10	0.7	0.5196	0.8804	0	1	0.1528	0.483	69.01%	30.0%
50		10	0.9	0.7819	1	0	1	0.1	0.3162	35.14%	10.0%
100		10	0.6	0.4072	0.7928	0	1	0.1633	0.5164	86.07%	40.0%

## Reproduction Summary

C-%	Control Type	Count	Mean	95% LCL	95% UCL	Min	Max	Std Err	Std Dev	CV%	%Effect
0	Dilution Water	10	26.1	23.22	28.98	5	31	2.438	7.709	29.54%	0.0%
6.25		10	12.5	7.319	17.68	0	31	4.387	13.87	111.0%	52.11%
12.5		10	17.3	12.55	22.05	0	32	4.025	12.73	73.57%	33.72%
25		10	6.1	3.463	8.737	0	18	2.233	7.062	115.8%	76.63%
50		10	13.3	11.05	15.55	0	22	1.904	6.019	45.26%	49.04%
100		10	1.6	0.6652	2.535	0	7	0.7916	2.503	156.5%	93.87%



# CETIS Summary Report

Report Date: 02 Jan-14 13:50 (p 2 of 2)  
Test Code: 1312-039 | 15-2517-8062

## Ceriodaphnia 7-d Survival and Reproduction Test

Rainier Environmental Laboratory

### 7d Survival Rate Detail

C-%	Control Type	Rep 1	Rep 2	Rep 3	Rep 4	Rep 5	Rep 6	Rep 7	Rep 8	Rep 9	Rep 10
0	Dilution Water	1	1	1	1	1	1	1	1	1	1
6.25		0	1	0	0	1	0	1	1	1	0
12.5		1	1	1	0	0	1	1	1	1	1
25		0	1	1	1	0	1	1	1	1	0
50		1	1	1	1	1	1	1	0	1	1
100		0	0	1	1	1	1	0	1	0	1

### Reproduction Detail

C-%	Control Type	Rep 1	Rep 2	Rep 3	Rep 4	Rep 5	Rep 6	Rep 7	Rep 8	Rep 9	Rep 10
0	Dilution Water	27	30	27	26	25	30	29	31	31	5
6.25		0	21	0	0	16	0	31	26	31	0
12.5		21	29	30	0	0	20	32	22	0	19
25		0	4	10	18	0	2	16	0	11	0
50		13	22	12	13	16	12	18	0	18	9
100		0	0	0	0	2	5	0	2	0	7

### 7d Survival Rate Binomials

C-%	Control Type	Rep 1	Rep 2	Rep 3	Rep 4	Rep 5	Rep 6	Rep 7	Rep 8	Rep 9	Rep 10
0	Dilution Water	1/1	1/1	1/1	1/1	1/1	1/1	1/1	1/1	1/1	1/1
6.25		0/1	1/1	0/1	0/1	1/1	0/1	1/1	1/1	1/1	0/1
12.5		1/1	1/1	1/1	0/1	0/1	1/1	1/1	1/1	1/1	1/1
25		0/1	1/1	1/1	1/1	0/1	1/1	1/1	1/1	1/1	0/1
50		1/1	1/1	1/1	1/1	1/1	1/1	1/1	0/1	1/1	1/1
100		0/1	0/1	1/1	1/1	1/1	1/1	0/1	1/1	0/1	1/1

Rainier Environmental  
Washington Laboratory

Client: Washington Beef  
Sample ID: WET  
Test No: 13-039  
Log-In#: 13-157 13-160

Initial and Final Chemistries

Seven Day Chronic Freshwater Bioassay  
Start Date & Time: 12/10/13 1530  
Stop Date & Time: 12/17/13 1500  
Test Species: Ceriodaphnia dubia  
13-168

Conc. or %	Days													
	0		1		2		3		4		5		6	
	init.	final	init.	final	init.	final	init.	final	init.	final	init.	final	init.	final
CON														
pH	7.45	7.47	7.44	7.35	7.37	7.25	7.42	7.38	7.32	7.41	7.34	7.45	7.47	7.51
DO (mg/l)	8.1	8.2	8.0	8.2	8.1	8.2	8.1	8.2	8.1	8.1	8.1	8.2	8.0	8.2
Cond. (µmhos-cm)	194	213	195	204	196	212	193	202	196	201	195	202	195	199
Temperature (°C)	24.6	25.8	25.1	25.1	24.9	25.4	24.5	25.2	24.2	25.4	25.1	25.5	24.5	25.2
Days														
6.25														
pH	7.41	7.62	7.42	7.58	7.37	7.34	7.42	7.44	7.31	7.45	7.34	7.45	7.45	7.55
DO (mg/l)	7.8	7.9	8.0	8.1	8.1	8.2	8.2	8.2	8.1	8.1	8.2	8.2	8.0	8.2
Cond. (µmhos-cm)	480	508	475	496	481	492	477	491	481	493	472	491	470	472
Temperature (°C)	24.7	25.6	25.1	25.6	24.9	25.4	24.7	25.3	24.2	25.4	25.0	25.5	24.3	25.3
Days														
12.5														
pH	7.38	7.66	7.35	7.71	7.35	7.44	7.39	7.51	7.31	7.57	7.32	7.55	7.41	7.62
DO (mg/l)	8.2	8.2	7.9	8.1	8.0	8.2	8.2	8.2	8.1	8.2	8.2	8.2	7.9	8.2
Cond. (µmhos-cm)	765	789	762	771	767	769	759	771	767	772	743	755	741	748
Temperature (°C)	24.5	25.7	25.2	25.5	24.8	25.4	24.4	25.3	24.5	25.4	25.0	25.5	24.3	25.3
Days														
25														
pH	7.33	7.77	7.32	7.82	7.33	7.52	7.35	7.59	7.25	7.69	7.28	7.67	7.35	7.68
DO (mg/l)	7.9	8.2	7.9	8.0	8.1	8.2	8.0	8.1	8.1	8.2	8.1	8.2	7.8	8.2
Cond. (µmhos-cm)	1308	1317	1311	1318	1309	1323	1298	1314	1305	1311	1284	1304	1279	1284
Temperature (°C)	24.5	25.7	25.3	25.5	24.8	25.4	24.5	25.5	25.1	25.4	25.1	25.5	24.2	25.3
Days														
50														
pH	7.25	7.84	7.21	7.89	7.23	7.71	7.22	7.73	7.22	7.81	7.18	7.78	7.27	7.80
DO (mg/l)	7.9	8.2	8.0	8.2	8.1	8.2	7.8	8.2	8.0	8.0	8.0	8.2	7.7	8.2
Cond. (µmhos-cm)	2350	2378	2347	2367	2355	2362	2335	2352	2350	2360	2297	2331	2291	2288
Temperature (°C)	25.1	25.5	25.4	25.5	25.1	25.4	24.6	25.3	24.8	25.4	25.3	25.5	24.4	25.2
Days														
100														
pH	7.18	7.90	7.13	7.99	7.17	7.91	7.15	7.94	7.16	7.92	7.19	7.89	7.22	7.94
DO (mg/l)	8.2	8.2	8.1	8.2	8.0	8.2	7.6	8.1	7.7	8.1	7.8	8.2	7.6	8.2
Cond. (µmhos-cm)	4470	4590	4450	4490	4460	4510	4440	4570	4490	4480	4180	4240	4150	4190
Temperature (°C)	24.9	25.6	25.7	25.5	25.2	25.4	25.2	25.3	24.7	25.4	25.8	25.5	24.7	25.3
Tech. Initials	ET	ET	ET	ET	ET	ET	ET	ET	ET	ET	ET	ET	ET	ET

Dilution Water Batch #: DMW 012  
Test Chamber: VWR

QA Check: ET

Sample Description: Brown  
Animal Source: In House Culture  
Comments: \_\_\_\_\_  
Date Received: \_\_\_\_\_ Date of Hatch: \_\_\_\_\_



Client/Sample ID: Washington Reef  
Test Number: 13/2-039

Ceriodaphnia 7-Day Chronic Survival and Reproduction

Start Date and Time: 12/10/13 1530  
Stop Date and Time: 12/17/13 1500

Rep	Conc.	Cont	Daily Reproduction								Day 6 Total	Third Brood
			1	2	3	4	5	6	7	8		
1	CON	54	—	—	—	5	—	9	13		14	27
2		51	—	—	—	5	—	11	14		16	30
3		41	—	—	—	4	—	10	13		13	27
4		33	—	—	—	3	—	10	13		13	26
5		21	—	—	—	4	—	8	13		13	25
6		53	—	—	—	4	—	11	15		15	30
7		15	—	—	—	4	—	10	15		14	29
8		16	—	—	—	3	—	11	17		14	31
9		52	—	—	—	4	—	12	15		16	31
10		7	—	—	—	3	—	—	2		3	5
Analyst												
Time												
Select #												
Rep	Conc.	Cont	1	2	3	4	5	6	7	8	Day 6 Total	Third Brood
1	6.25	3	—	—	X	—	—	8	8		0	0
2		3	—	—	—	5	—	—	8		13	21
3		46	—	—	X	—	—	—	—		0	0
4		42	—	—	X	—	—	—	—		0	0
5		32	—	—	—	4	—	12	—		0	16
6		30	—	X	—	—	—	—	—		0	0
7		24	—	—	—	5	12	—	14		17	31
8		25	—	—	—	5	6	—	15		11	26
9		59	—	—	—	4	—	—	16		15	31
10		44	—	—	X	—	—	—	—		0	0

Rep	Conc.	Cont	Daily Reproduction								Day 6 Total	Third Brood
			1	2	3	4	5	6	7	8		
1	25	14	—	—	X	—	—	2	—		0	0
2		35	—	—	—	2	—	2	—		4	4
3		55	—	—	—	2	—	5	—		7	10
4		45	—	—	—	2	—	10	3		15	18
5		49	X	—	—	—	—	—	3		0	0
6		40	—	—	—	—	—	2	—		2	2
7		1	—	—	—	2	—	6	8		8	16
8		60	—	—	—	—	—	—	—		8	16
9		56	—	—	—	—	—	—	—		8	16
10		12	—	—	X	—	—	—	—		0	0

Rep	Conc.	Cont	Daily Reproduction								Day 6 Total	Third Brood
			1	2	3	4	5	6	7	8		
1	50	4	—	—	—	—	—	4	9		4	13
2		19	—	—	—	—	—	8	14		8	22
3		50	—	—	—	—	—	6	6		8	22
4		22	—	—	—	—	—	9	4		9	13
5		52	—	—	—	—	—	8	8		8	16
6		36	—	—	—	—	—	2	10		2	16
7		17	—	—	—	—	—	6	12		6	19
8		9	—	X	—	—	—	—	—		0	0
9		43	—	—	—	—	—	6	12		0	0
10		47	—	—	—	—	—	—	12		0	0

Rep	Conc.	Cont	Daily Reproduction								Day 6 Total	Third Brood
			1	2	3	4	5	6	7	8		
1	12.5	26	—	—	—	5	—	4	12		9	21
2		37	—	—	—	4	—	11	14		15	29
3		29	—	—	—	3	—	8	19		11	30
4		57	—	—	X	—	—	—	—		0	0
5		39	X	—	—	4	—	4	12		0	0
6		6	—	—	—	8	—	11	12		8	22
7		10	—	—	—	4	—	11	13		7	22
8		13	—	—	—	4	—	—	15		7	22
9		20	—	—	—	3	—	—	—		0	0
10		28	—	—	—	3	—	6	10		0	0

Rep	Conc.	Cont	Daily Reproduction								Day 6 Total	Third Brood
			1	2	3	4	5	6	7	8		
1	100	11	—	—	X	—	—	—	—		0	0
2		2	—	—	X	—	—	—	—		0	0
3		19	—	—	—	—	—	—	—		0	0
4		34	—	—	—	—	—	—	—		0	0
5		31	—	—	—	—	—	—	—		0	0
6		27	—	—	—	—	—	—	—		0	0
7		53	X	—	—	—	—	—	—		0	0
8		38	—	—	—	—	—	2	—		0	0
9		44	—	X	—	—	—	—	—		0	0
10		44	—	—	—	—	2	5	—		0	0

Comments: X=mortality

QA 24



**Appendix B**  
**Sample Check-In Sheets**

Rainier Environmental  
5013 Pacific Hwy East, Ste. 20  
Tacoma, WA 98424

Sample Check-In Information

Client: Washington Bay

Tests Performed: cd-c  
Test ID No(s): 1312-039

Sample Description:  
Brown

Sample ID:	WET	WET	FE effluent CO2 WET
Log-in No. (10-xxxx):	13-157	13-160	13-168
Sample Collection Date & Time:	12/9/13 720	12/11/13 725	12/13/13 740
Sample Receipt Date & Time:	12/10/13 1030	12/12/13 1400	12/14/13 1100
Check-in Temperature (°C)	1.0	2.4	2.4
Temperature OK?	<input checked="" type="radio"/> Y <input type="radio"/> N	<input checked="" type="radio"/> Y <input type="radio"/> N	<input checked="" type="radio"/> Y <input type="radio"/> N
DO (mg/L)	6.7	7.0	5.4
pH (units)	7.47	7.35	7.30
Conductivity (µS/cm)	4610	4390	4090
Salinity (ppt)	2.3	2.3	2.0
Tit. Vol / Sam. Vol. / Alkalinity (mg/L)*	42.1 25.1 168	40.1 25.1 160	44.1 25.1 176
Tit. Vol. / Sam. Vol. / Hardness (mg/L)*	20.1 25.1 90	19.1 25.1 76	19.1 25.1 76
Total Chlorine (mg/L)	2003	2003	2003
Total Ammonia (mg/L)	21.0	21.0	21.0
Technician Initials	gt	gt	gt

\* = mg/L as CaCO<sub>3</sub>; \* = Measured for freshwater samples only, NA = Not Applicable,  
NM = Not Measured

Freshwater Tests:

Control/Dilution Water Source: test type: cd-c 8.2 (DMW) -0.2 ART SW Other: 64 Alkalinity: 64 Hardness: 88  
Control/Dilution Water Source: test type: 8.2 (DMW) MH-W Other: 64 Alkalinity: 64 Hardness: 88  
Additional Control? Y N = 64 Alkalinity: 64 Hardness: 88

Marine Tests:

Control/Dilution Water Source: test type: ART SW NAT SW Alkalinity: 64 Salinity: 88  
Control/Dilution Water Source: test type: ART SW NAT SW Alkalinity: 64 Salinity: 88  
Additional Control? Y N = 64 Alkalinity: 64 Salinity: 88  
Sample Salted w/ artificial salt? Y N If yes, what ppt? 64 test type: 64  
Sample salted w/brine? Y N If yes, what ppt? 64 test type: 64

Comments: Temperature for grab sample must be 0-20°C if received within 1 hour of collection time, 0-12°C if effluent received within 4 hours of collection time, and 0-6°C for all other samples.

COC Complete? Y or N  
1 Y 2 Y 3 Y

Filtration? Y (N)  
Pore Size:                       
Organisms or Debris                     

Aeration? Y (N)  
Length of Time:                       
Final DO:                       
Final pH:                     

Hardness Adjustment? Y (N)  
If adjusted, please see worksheet for details.

Sub-samples for additional chemistry:

QC Check: gt

**Appendix C**  
**Reference Toxicant Test**



**Appendix D**  
**Chain-of-Custody Forms**



ier  
ONMENT

**Washington**  
5013 Pacific Highway East, Suite 20 Fife,  
WA 98424  
Phone 253 922 8898

## Chain of Custody

Date 12/19/13 Page 1 of 1

[illegible]



**Washington**  
5013 Pacific Highway East, Suite 20 Fife,  
WA 98424  
Phone 253.922.8898

Date 12/11/13 Page 1 of 1

[illegible]



Sample Collection By:

ANALYSES REQUIRED

Date \_\_\_\_\_ Page \_\_\_\_\_ of \_\_\_\_\_

**Report to:**

Company Washington Beef LLC  
Address 201 Elmwood Rd.  
City/State/zip Toppenish, WA 98948  
Contact Sherry Byers-Eddy  
Phone 509-452-6534  
Email Sherry.Byers@abfoodusa

**Invoice To:**

Company Washington Beef LLC  
Address 201 Elmwood Rd.  
City/State/zip Toppenish, WA 98948  
Contact Sherry Byers-Eddy  
Phone 509-452-6534  
Email Sherry.Byers@abfoodusa

Receipt Temperature (°C)

SAMPLE ID	DATE	TIME	MATRIX	CONTAINER TYPE	NO. OF CONTAINERS	COMMENTS
1 F.E. Outfall 002-WET	12/13/13	7:40 am	WASTE water	containers	1	WET
2						
3						
4						
5						
6						
7						
8						
9						
10						

  

PROJECT INFORMATION		SAMPLE RECEIPT		RELINQUISHED BY (CLIENT)		RELINQUISHED BY (COURIER)	
Client:		Total No. of Containers	1	(Signature)	<u>Mike Hernandez</u>	(Signature)	
PO No.:		Received Good Condition?	Y	(Printed Name)	<u>Mike Hernandez</u>	(Printed Name)	
Shipped Via:	<u>UPS</u>	Matches Test Schedule?	Y	(Company)	<u>WA. Beef LLC.</u>	(Company)	
SPECIAL INSTRUCTIONS/COMMENTS:				RECEIVED BY (COURIER)		RECEIVED BY (LABORATORY)	
				(Signature)		(Signature)	<u>Eric Tolleson</u>
				(Printed Name)		(Printed Name)	<u>ERIC TOLLESON</u>
				(Date)		(Date)	<u>12/14/13</u>
				(Log in #)		(Log in #)	<u>13-168</u>